

FACTOR XI

Targeting factor XI and factor XIa to prevent thrombosis

David Gailani¹ and Andras Gruber²

¹The Department of Pathology, Microbiology and Immunology, Vanderbilt University Medical Center, Nashville, TN; and ²Aronora, Inc, Portland, OR

Direct oral anticoagulants (DOACs) that inhibit the coagulation proteases thrombin or factor Xa (FXa) have replaced warfarin and other vitamin K antagonists (VKAs) for most indications requiring long-term anticoagulation. In many clinical situations, DOACs are as effective as VKAs, cause less bleeding, and do not require laboratory monitoring. However, because DOACs target proteases that are required for hemostasis, their use increases the risk of serious bleeding. Concerns over therapy-related bleeding undoubtedly contribute to undertreatment of many patients who would benefit from anticoagulation therapy. There is considerable interest in the plasma zymogen factor XI (FXI) and its protease form factor XIa (FXIa) as drug targets for treating and preventing thrombosis. Laboratory and epidemiologic studies support the conclusion

that FXI contributes to venous and arterial thrombosis. Based on 70 years of clinical observations of patients lacking FXI, it is anticipated that drugs targeting this protein will cause less severe bleeding than warfarin or DOACs. In phase 2 studies, drugs that inhibit FXI or FXIa prevent venous thromboembolism after total knee arthroplasty as well as, or better than, low molecular weight heparin. Patients with heart disease on FXI or FXIa inhibitors experienced less bleeding than patients taking DOACs. Based on these early results, phase 3 trials have been initiated that compare drugs targeting FXI and FXIa to standard treatments or placebo. Here, we review the contributions of FXI to normal and abnormal coagulation and discuss results from preclinical, nonclinical, and clinical studies of FXI and FXIa inhibitors.

Introduction

In 1977, in an edition of the *British Medical Bulletin* devoted to the pathology of blood coagulation, the Oxford University researcher R. Gwynn Macfarlane described thrombosis as “hemostasis in the wrong place.”¹ Macfarlane, who was first to propose that the coagulation protease thrombin (factor IIa [FIIa]) is formed by a cascading sequence of plasma enzymatic reactions,² suggested that “the key to the problem of thrombosis lies in a deeper understanding of the negative aspects of hemostasis.”¹ A conclusion readily drawn from this intuitive premise is that thrombosis can be treated or prevented by inhibiting processes required for the normal response to injury. This principle has guided development of anticoagulation therapy with remarkable success. However, bleeding associated with vitamin K antagonists (VKAs) and, more recently, with direct oral anticoagulants (DOACs), seems inevitable with this approach. Anticoagulation-related bleeding causes significant morbidity and mortality, limits the intensity of therapy that can be applied, and places restrictions on who can receive treatment. This reality is the driving force behind efforts to develop drugs that separate antithrombotic activities from deleterious effects on hemostasis.^{3,4} Here, we discuss the rationale for inhibiting the plasma protein factor (F) XI or its protease form FXIa to improve the safety of antithrombotic therapy.

Thrombin generation and anticoagulation

The protease thrombin is central to the host response to injury, catalyzing fibrinogen conversion to fibrin, stimulating platelets and cells by cleavage of protease activated receptors, and activating clotting factors.^{5,6} In current models of plasma coagulation, thrombin is the product of a set of reactions involving vitamin K-dependent proteases and their cofactors (indicated by yellow arrows in [Figure 1A](#)) that are initiated when FVIIa forms a complex with the cofactor tissue factor, which is expressed on surfaces of cells at an injury site.⁷⁻¹⁰ Studies with genetically altered mice indicate that absence of any component of this core system is either incompatible with life (prothrombin, FV, FVII, FX, and tissue factor), or causes severe bleeding (FVIII and FIX).¹¹ For the past 80 years, components of this vital mechanism have been the primary targets of anticoagulation therapy.

Current anticoagulants inhibit the activity of thrombin (dabigatran, bivalirudin, and argatroban), FXa (rivaroxaban, apixaban, edoxaban, and fondaparinux), or both proteases (unfractionated heparin, low molecular weight heparin, and danaparoid) or reduce plasma concentrations of their precursors (VKAs). Because these proteases are required for hemostasis, drugs

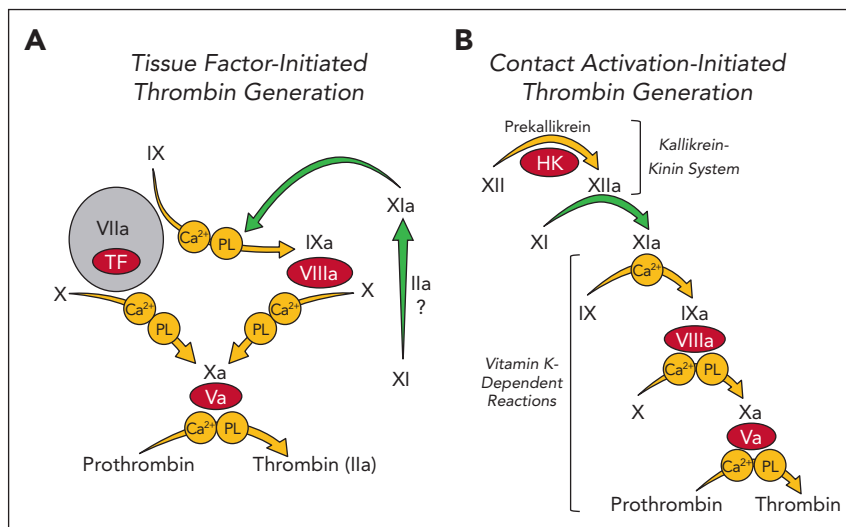


Figure 1. Models of thrombin generation. Shown are major enzymatic reactions (represented by arrows) during plasma coagulation. In each reaction a plasma zymogen (black Roman numerals) is converted to an active protease (Roman numerals followed by “a”). Coagulation cofactors are shown as red ovals. Requirements for calcium ions (Ca^{2+}) or phospholipids (PLs) for specific reactions are indicated within circles. Green arrows indicate reactions involving factor XI (XI) or factor XIa (XIa). (A) Tissue factor (TF)-initiated thrombin generation. Coagulation is initiated when FVIIa binds to TF that is exposed to blood at a wound site (gray circle). The FVIIa-TF complex converts FX to FXa leading to initial thrombin generation. FVIIa-TF also converts FIX to FIXa, which sustains thrombin generation by activating additional FX. In this system FXIa serves a limited role, generating some FIXa. FXI may be activated by thrombin or other unknown proteases (?) in this system. Its activation is not dependent on FXII. (B) Contact activation-initiated thrombin generation. The process is initiated by exposure of plasma to a surface (typically carrying a negative charge), which triggers FXII activation by a process called contact activation. This sets off a series (cascade) of enzymatic reactions culminating in conversion of prothrombin to thrombin. Here, FXI is a component of the contact activation process that triggers coagulation and is a major contributor to thrombin generation.

targeting them can cause bleeding that increases with anti-coagulation intensity. From the standpoint of avoiding bleeding, DOACs (which target 1 protease) are superior to warfarin (which reduces FII, FVII, FIX, and FX plasma concentrations).¹²⁻¹⁴ However, the annual risk of serious bleeding in patients with atrial fibrillation on DOACs to prevent stroke is as high as 3%, with intracranial bleeding occurring in 0.3% to 0.5%.¹² Bleeding concerns likely contribute to one-third of patients with atrial fibrillation not receiving anticoagulation and to underanticoagulation in many who are treated.¹⁵⁻¹⁷ According to a meta-analysis by Khan et al of patients receiving DOACs for venous thromboembolism (VTE), the rate of major bleeding events was 1.12 (confidence interval [CI], 0.72-1.62) per 100 patient years, with a case-fatality rate of 9.7% (CI, 3.3%-19.2%).¹⁸ Rates were even higher for patients with reduced kidney function, with a bleeding history, on antiplatelet agents, or aged >65 years.¹⁸ In addition, the safety advantage that DOACs have over VKAs may partly reflect less intense anti-coagulation and, thus, a weaker antithrombotic effect. This may partly explain why DOACs are less effective than warfarin for patients with mechanical heart valves,^{19,20} antiphospholipid antibody syndrome,^{21,22} and atrial fibrillation caused by rheumatic heart disease.²³

FXI in thrombin generation and hemostasis

FXI is the zymogen of the protease FXIa.²⁴ Its properties are discussed in the accompanying paper by Moellmer et al.²⁵ FXI was first identified as a constituent of plasma missing in individuals with mild bleeding symptoms but with a defect in *in vitro* clotting similar to the one in the severe bleeding disorders hemophilia A and B (deficiency of FVIII or FIX, respectively).²⁶ In the activated partial thromboplastin time (aPTT)

assay, thrombin generation is initiated when FXII is converted to the protease FXIIa by a process called contact activation (Figure 1B).^{27,28} FXIIa converts FXI to FXIa which then activates FIX. Consequently, the aPTT is prolonged in plasma lacking FXI.²⁸ Indeed, FXI-deficient plasma usually has a longer aPTT than plasma lacking FVIII or FIX. Despite this, bleeding is milder in patients who are FXI deficient than in those with hemophilia and follows a different pattern (refer to the accompanying paper by Barg et al).²⁹⁻³¹ Plasma FXI levels correlate poorly with bleeding propensity,^{28,30,32,33} and many individuals with FXI deficiency do not experience abnormal hemostasis. Furthermore, persons lacking FXII, the activator of FXI in the aPTT, do not have a hemostatic defect.²⁸ From this, it is reasonable to conclude that the role of FXI in hemostasis differs from its role in the aPTT assay.

In contrast to its major contribution in the aPTT, FXI serves a minor role in tissue factor-initiated models of thrombin generation, in which it supplements FIXa produced by the FVIIa-tissue factor complex (Figure 1A).^{6,8,10} Clinical observations suggest that FXIa activity is most important for hemostasis when injury involves tissues with robust fibrinolytic activity, such as the nasopharynx, mouth, and urinary tract.²⁸⁻³³ Because FXII deficiency does not cause abnormal bleeding, it is likely that FXI is activated by proteases other than FXIIa during hemostasis. For example, thrombin converts FXI to FXIa (Figure 1A).^{24,25,34-36} Other proteases may activate it as well.

Evidence from animal studies implicating FXI in thrombosis

The limited role of FXI in hemostasis might suggest that its impact on thrombosis would also be small. However, data from animal models indicate that FXI is an important contributor to

thrombosis. In 2003, Gruber and Hanson described the effects of a polyclonal anti-FXI immunoglobulin G (IgG) on experimentally induced thrombosis in baboons.³⁷ Although the antibody did not prevent clots from forming on surfaces of Dacron or tissue factor–coated Teflon grafts inserted into arteriovenous shunts, it greatly reduced clot growth into the graft lumen. Intraluminal growth is necessary for a thrombus to occlude a vessel but is likely less important for hemostasis, during which clots form primarily within walls of damaged vessels and in extravascular spaces. In this study, the antithrombotic effect of FXI inhibition was comparable with that of unfractionated heparin. However, unlike heparin, FXI inhibition did not prolong the bleeding time. The authors concluded that targeting FXI could be antithrombotic without the profound impairment of hemostasis caused by anticoagulants targeting other coagulation factors.³⁷ Subsequent studies with monoclonal IgGs supported the premise that thrombus growth requires FXI in the vascular graft model^{38,39} and suggest that as a thrombus expands away from the vessel wall containing the stimulus for clotting, FXI sustains thrombin generation to drive clot expansion.

FXI also supports thrombosis in rodents.^{40–44} In mice in which carotid artery occlusion is induced by injuring the vessel with ferric chloride (FeCl₃), an infusion of unfractionated heparin produces an antithrombotic effect that is reflected in the higher FeCl₃ concentration needed to occlude the vessel (Figure 2A, compare red and blue bars).⁴¹ Comparable resistance to thrombosis occurs in mice lacking FIX, a model of hemophilia B (Figure 2A, orange bars).^{41,45} For both heparin infusion and FIX deficiency, the antithrombotic effect is accompanied by severe bleeding after tail transection (Figure 2B), consistent with disrupted hemostasis. FXI-deficient mice are as resistant to FeCl₃-induced carotid artery occlusion as FIX-deficient mice (Figure 2A, compare orange and green bars), but their tail bleeding times are comparable with those of normal mice (Figure 2B).⁴¹ Indeed, FXI deficiency or FXIa inhibition has not

caused major hemostatic abnormalities in any nonhuman species in which it has been tested.

Inhibition or deficiency of FXII also reduces thrombus formation in animal models. The effect is most striking in mice^{46–48} but is observed in primates.⁴⁹ Because FXII deficiency does not cause abnormal bleeding, FXII inhibition could be an even safer antithrombotic strategy than FXI inhibition. However, the acute vessel injuries used to induce thrombosis in healthy animals may not mirror the biology of disorders, such as VTE, myocardial infarction (MI), or ischemic stroke, in which clots form in the setting of chronic vascular disease. Data from population studies have been important for assessing the relative contributions of FXI and FXII to thrombosis in humans.

Data implicating FXI in thrombosis in humans

In 2000, Meijers et al described an association between plasma FXI and VTE risk in enrollees in the Leiden Thrombophilia Study.⁵⁰ Participants with FXI antigen concentrations >90th percentile had a more than twofold increased risk compared with the rest of the group, independent of other known genetic risk factors. Subsequent studies reported correlations between plasma FXI levels and incidences of VTE,^{51–55} MI^{54,56–58}, and ischemic stroke.^{54,58–60} VTE and ischemic stroke are also less frequent in individuals with FXI deficiency than in the general population.^{61,62} FXI deficiency is particularly prevalent in people of Ashkenazi Jewish ancestry.^{28–30} In 2017, Preis et al presented results from a historical cohort study of >10 000 patients in a health care system in Israel: 690 with mild FXI deficiency (activity 30%–50% of normal) and 542 with moderate-to-severe deficiency (≤30% of normal).⁵⁴ The adjusted hazard ratio of VTE for those with FXI deficiency was 0.26 (95% CI, 0.08–0.84). Arterial events (MI, stroke, and transient ischemic attacks) were also lower in patients with mild (0.52; 95% CI 0.31–0.87) or

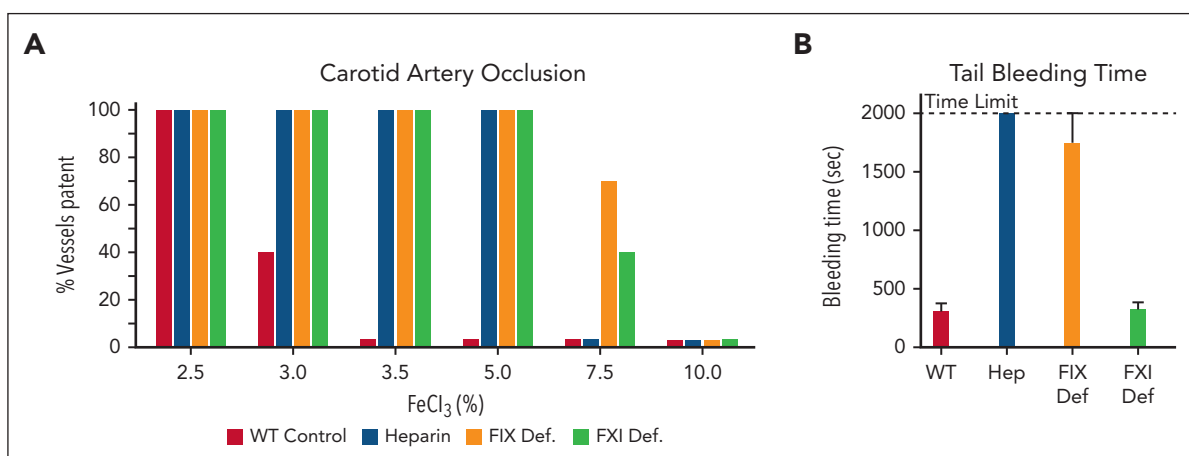


Figure 2. Separating an antithrombotic effect from an antihemostatic effect. (A) Ferric chloride–induced carotid artery occlusion model. The carotid artery is briefly exposed to a pad saturated with FeCl₃, and blood flow through the vessel is monitored for 30 minutes. Each bar represents a group of 10 mice, and the bar height indicates the percent of each group with patent blood vessels at the end of the experiment. Note that all wild-type mice (red bars) have occluded arteries with 3.5% FeCl₃, whereas mice treated with heparin (blue bars), or mice deficient in FIX (orange bars) or FXI (green bars) require higher concentrations of FeCl₃ to induce vessel occlusion. (B) Tail bleeding times. Wild-type mice treated with vehicle (red bar) or heparin (blue bar), or mice lacking FIX (orange bar), or FXI (green bar) underwent removal of the tail tip. The bleeding tail was immersed in normal saline at 37°C, and time to cessation of bleeding was recorded. Results are the mean bleeding times for groups of 10 mice ± 1 standard deviation. If bleeding did not stop by the 2000-second time point, the tail was cauterized.

moderate-to-severe (0.57; 95% CI, 0.35-0.93) FXI deficiency than for the general population.

FXI vs FXII as an antithrombotic target

Over the past decade, efforts to develop safer antithrombotic strategies have focused more on FXI than on FXII for 2 reasons. First, in contrast to the epidemiologic data for FXI, evidence that FXII influences VTE, MI, or stroke risk in humans is weak. Most recent studies show a negative correlation,^{55,57,63,64} and 2 reported inverse relationships for MI and cardiovascular death.^{57,64} Second, FXIIa is probably not the only FXI activator in vivo. The FVIIa-tissue factor complex is an important trigger for venous and arterial thrombus formation,⁹ and FXI is activated independently of FXIIa during tissue factor-initiated coagulation (Figure 1A).^{34-36,65} Based on these considerations, drugs targeting FXI or FXIa (hereafter referred to collectively as FXI/XIa inhibitors) may have advantages over drugs directed at FXII in some clinical situations.

This is not meant to imply that FXII inhibition is not a useful antithrombotic strategy. FXII triggers surface-mediated thrombin generation, which contributes to thrombosis when blood is exposed to medical devices during renal dialysis, cardiopulmonary bypass, or extracorporeal membrane oxygenation.⁶⁶⁻⁶⁸ Artificial heart valves, ventricular assist devices, and venous catheters are also prothrombotic, at least partly through induction of contact activation.^{67,69,70} Inhibiting FXIIa could reduce surface-induced thrombus formation with little, if any, impact on hemostasis.

FXI inhibition and bleeding

Because FXI deficiency causes a mild bleeding diathesis, it is anticipated that FXIa inhibitors will cause less major bleeding than DOACs. However, FXI deficiency can compromise hemostasis, and we should anticipate treatment-related bleeding in

some patients on these drugs. Excessive bleeding after trauma, particularly to the nose, mouth, or genitourinary tract, may occur. Although unprovoked bleeding is likely to be rare, heavy menstrual bleeding may occur. A recent analysis by de Jong et al of women aged between 18 and 50 years on anti-coagulation for acute VTE (87% on DOACs) found that 2 of 3 experienced abnormal uterine bleeding.⁷¹ Heavy menstrual bleeding is more common in women who are FXI deficient than in the general population,⁷² suggesting that it may be induced or aggravated by FXI/XIa inhibitors.

Perhaps most important from the standpoint of comparison to VKAs and DOACs is that intracranial and gastrointestinal bleeding are not prominent features of severe FXI deficiency. Thus, there may be relatively few life-threatening bleeds in patients administered FXI/XIa inhibitors, regardless of the overall impact on bleeding incidence. If this is the case, drugs targeting FXI or FXIa could be used in ways that are not practical with thrombin or FXa inhibitors. For example, drug dose could be pushed to abolish FXI or FXIa activity to maximize an antithrombotic effect. It may also be possible to use agents with long half-lives that can be administered every few weeks.

Drugs targeting FXI and FXIa

Table 1 lists drugs that reduce plasma FXI or inhibit FXI or FXIa that have undergone phase 2 testing. Long-acting agents include DNA antisense oligonucleotides (ASOs) that reduce plasma FXI antigen by promoting degradation of FXI messenger RNA in the liver (lonis-FXI Rx),^{73,74} antibodies that bind FXI and interfere with its activation and activity (abelacimab and xisomab [gruticibart]),^{39,75,76} and antibodies that block the FXIa active site (osocimab).⁷⁷ The small molecule oral FXIa active site inhibitors milvexian and asundexian have half-lives more comparable with those of current DOACs.^{78,79} FXI/XIa inhibitors have been tested for efficacy and/or safety as VTE

Table 1. Drugs targeting FXI and FXIa studied in phase 2 trials

Drug name	Drug description	Mechanism of action	Administration	Duration of effect
lonis FXI _{Rx} (BAY-2306001)	DNA antisense oligonucleotide with complementary sequence to FXI messenger RNA	Reduces synthesis of FXI in hepatocytes	Subcutaneous or IV	Effect may persist for weeks to months after discontinuation
Osocimab	Monoclonal IgG that binds to the catalytic domain of FXIa	Inhibits FXIa catalytic activity through allosteric effects	Subcutaneous or IV	Days to weeks depending on dose. Mean elimination half-life, 30-44 days.
Abelacimab	Monoclonal IgG that binds to the catalytic domain of FXI and FXIa	Binds FXI preventing its activation. It also neutralizes FXIa activity	Subcutaneous or IV	Mean elimination half-life, 25-30 days. The IgG becomes saturated with FXI over time.
Xisomab (gruticibart)	Monoclonal IgG that binds to the apple 2 domain of FXI and FXIa	Interferes with FXI activation by FXIIa	Subcutaneous or IV	Half-life, 121.5 h with single dose of 5 mg/kg
Milvexian	Small molecule inhibitor of the FXIa catalytic active site	Inhibits FXIa catalytic activity by active site binding	Oral	Half-life, 11.4 to 18.1 h when administered daily
Asundexian	Small molecule inhibitor of the FXIa catalytic active site	Inhibits FXIa catalytic activity by active site binding	Oral	Dose dependent terminal half-life of 14.2 to 17.4 h

prophylaxis, for secondary stroke prevention, and in patients with heart disease or renal failure. Several excellent reviews of completed and ongoing clinical trials have been published recently.^{4,80-83} Here, we focus on findings that illustrate potential strengths and weaknesses of FXI/XIa inhibitors.

FXI or FXIa inhibition for VTE prevention

Anticoagulants are often tested first as VTE prophylaxis for patients undergoing total knee arthroplasty (TKA). In the absence of prophylaxis, 40% to 60% of patients undergoing TKA are expected to form clots in the involved leg that are detectable with venography. Results for 4 phase 2 trials of FXI or FXIa reduction or inhibition in patients undergoing TKA are available (Table 2).^{74,75,77,84} Their effects on clot formation determined by venography 8 to 14 days after surgery were compared with those of enoxaparin (40 mg/d) administered for at least 10 days. Bleeding was the primary safety end point, but the trials were of insufficient size to identify significant differences in bleeding between treatment arms.

Bueller et al investigated the anti-FXI ASO Ionis-FXI Rx (FXI-ASO TKA trial).⁷⁴ Because this drug takes 3 weeks to achieve stable reduction of plasma FXI, patients started treatment 36 days before surgery and were under the full effect of the drug

during surgery. Average plasma FXI levels were reduced to 38% and 20% of normal in patients on 200 or 300 mg doses of ASO, respectively. The incidence of venous clot formation was significantly lower in patients receiving 300 mg ASO (4%) than in those receiving enoxaparin (30%) or 200 mg ASO (27%), indicating a dose response for the ASO, with a near maximal effect at a FXI activity of ~20% of normal.

In the FOXTROT trial, Weitz et al compared single doses of osocimab to enoxaparin or apixaban (2.5 mg twice daily).⁷⁷ Osocimab blocks the FXIa active site, and a dose of 1.8 mg/kg increased the aPTT to ~1.6× normal. This is comparable with the effect of 300-mg Ionis-FXI Rx in the ASO trial.⁷⁴ Venous clots occurred in 11.3% and 17.9% of patients receiving 1.8 mg/kg osocimab before or after surgery, respectively, in 26.3% with enoxaparin, and in 14.5% with apixaban.

In the ANT-005 TKA trial, Verhamme et al compared single 30, 75, or 150 mg doses of the IgG abelacimab administered within 8 hours after surgery.⁷⁵ FXI bound to abelacimab is locked in a zymogen conformation. Because the normal plasma FXI concentration is between 15 and 45 nM, each abelacimab dose was sufficient to saturate FXI in most patients. As a result, peak aPTTs for all doses were similar and higher (~1.9× normal) than those in the ASO or FOXTROT studies. Clots occurred in 13%, 5%, and 4% of the patients on 30, 75, and 150 mg abelacimab, respectively, and in 22% on enoxaparin.

Table 2. Trials of drugs targeting FXI(a) for patients undergoing TKA

Clinical trial (drug)	Dosing	Primary efficacy end point	Primary safety end point	Incidence of venous thrombosis	Incidence of bleeding
FXI-ASO TKA (Ionis FXI Rx)	9 subcutaneous doses starting 36 days before op. Last dose 3 days after op	VTE by bilateral venography 8-12 d after op or symptomatic VTE	Major or clinically relevant nonmajor bleeding out to 100 d after op	Ionis FXI _{Rx} 200 mg (27%)* Ionis FXI _{Rx} 300 mg (4%) [†] Enoxaparin (30%)	Major or CRNM bleeding: Ionis FXI _{Rx} 200 mg (3%) and 300 mg (3%) Enoxaparin (8%)
FOXTROT (osocimab)	Single IV dose the day after surgery or just before surgery	VTE by bilateral venography 10-13 d after op or symptomatic VTE	Major or clinically relevant nonmajor bleeding out to 10-13 d after op	Post-op osocimab (mg/kg): 0.3 (23.7%), 0.6 (15.7%)*, 1.2 (16.5)*, 1.8 (17.9%)* Pre-op osocimab (mg/kg): 0.3 (29.9%) and 1.8 (11.3%) [†] Enoxaparin (26.3%) Apixaban (14.5%)	Major or CRNM bleeding: Osocimab after op (0%-3%) Osocimab before op (1.9%-5.9%) Enoxaparin (5.9%) Apixaban (2%)
ANT-005 TKA (abelacimab)	Single IV dose 4-8 h after op	VTE by venography of involved leg 8-12 d after op or symptomatic VTE	Major or clinically relevant nonmajor bleeding out to 30 d after op	Abelacimab: 30 mg (13%)*, 75 mg (5%) [†] , and 150 mg (4%) [†] Enoxaparin (22%)	Major or CRNM bleeding: Abelacimab 30 mg (2%), 75 mg (2%), and 150 mg (0%) Enoxaparin (0%)
AXIOMATIC-TKR (milvexian)	Once-daily or BID oral dosing starting 12-24 h after op, for 10-14 d	VTE by venography of involved leg 10-14 d after op, symptomatic VTE or death	Major, clinically relevant nonmajor, and minimal bleeding out to 6 wks after op	Milvexian BID: 25 mg (21%), 50 mg (11%) [‡] , 100 mg (9%) [‡] , and 200 mg (8%) [‡] Milvexian once per day: 25 mg (25%), 50 mg (24%), and 200 mg (7%) [‡] Enoxaparin (21%)	Major or CRNM bleeding: Milvexian (1%) and enoxaparin (2%) Any bleeding: Milvexian (4%) and enoxaparin (4%)

BID, twice daily; CRNM, clinically relevant nonmajor bleeding; op, operation.

*Noninferior compared with enoxaparin.

[†]Superior compared with enoxaparin.

[‡]Significantly lower than with enoxaparin.

In the AXIOMATIC-TKA trial, Weitz et al studied the oral FXIa inhibitor milvexian administered for 10 to 14 days, starting from 12 to 24 hours after TKA.⁸⁴ Patients receiving a daily dose of milvexian of ≥ 100 mg (administered once or twice daily) had significantly fewer clots than patients on enoxaparin (7%-11% vs 21%, respectively). aPTT ratios for patients receiving ≥ 100 mg milvexian per day were at least twofold above baseline by day 4 of therapy.

These studies showed that reducing FXI or FXIa activity significantly reduces venous clot formation after TKA compared with enoxaparin and support observations that inherited FXI deficiency decreases VTE risk.⁵⁰⁻⁵⁵ Severe bleeding was uncommon (Table 2) despite some patients having low FXI or FXIa activity during surgery (AXIOMATIC-TKA and FOXTROT trials)^{74,77} or nearly undetectable activity after operation (ANT-005 TKA trial).⁷⁵ Although severe bleeding is not common after TKA, the results support the impression that FXI or FXIa inhibition is safe in this setting.

Targeting FXIa for secondary prevention of ischemic stroke

Two trials compared the oral FXIa inhibitors asundexian (PACIFIC-STROKE)⁸⁵ and milvexian (AXIOMATIC-SSP)⁸⁶ with placebo for patients with recent acute stroke who were on antiplatelet therapy with aspirin and/or clopidogrel (Table 3). The primary efficacy outcomes were recurrent symptomatic ischemic stroke or magnetic resonance imaging (MRI)-detected covert infarct. As with the TKA trials, the stroke prevention trials were not large enough to establish significant differences in bleeding. Shoamanesh et al reported results for the PACIFIC-STROKE trial, in which patients were randomized to receive asundexian (10, 20, or 50 mg/d) or placebo.⁸⁵ Over 26 weeks, the primary efficacy outcome occurred at similar rates in the asundexian (19%-22%) and placebo (19%) arms. International Society on Thrombosis and Haemostasis major or clinically relevant nonmajor bleeding occurred in 3% to 4% of patients administered asundexian and 2% on placebo. In the AXIOMATIC-SSP trial, Sharma et al assigned patients to milvexian (25-200 mg, once or twice daily) or placebo for 90

days.⁸⁶ The primary safety end point was major bleeding (Bleeding Academic Research Consortium–modified type 3 and 5 bleeding). There were no differences between treatment arms for the primary efficacy or safety endpoints (Table 3).

There are several issues to consider regarding the negative results of these trials. Obviously, although FXI deficiency appears to reduce risk of initial stroke in population studies,^{54,58-60} this does not mean that FXIa inhibition will reduce stroke recurrence. Also, FXIa inhibition may not add to the protective effect provided by dual antiplatelet therapy. Issues around study design also need consideration. The doses of FXIa inhibitors might have been insufficient to affect stroke recurrence. Although the higher doses of milvexian used in the AXIOMATIC-SSP trial reduced VTE in the AXIOMATIC-TKA trial,⁸⁴ the doses of asundexian used in the PACIFIC-STROKE trial have not been shown to reduce thrombus formation in humans. Regarding the primary efficacy end points, approximately two-thirds of events in both trials were asymptomatic lacunar infarcts detected by MRI. FXI or FXIa inhibition may not prevent these common poststroke events. In a post-hoc analysis, symptomatic recurrent ischemic strokes were reduced in patients receiving the FXIa inhibitor in the PACIFIC-STROKE and AXIOMATIC-SSP trials. The OCEANIC-STROKE and LIBREXIA-STROKE trials (Table 4) will continue the evaluation of FXIa inhibition in preventing recurrent stroke.

Bleeding in patients with heart disease on FXI/XIa inhibitors

Three phase 2 trials were designed to address the impact of FXI or FXIa inhibition on hemostasis in patients with heart disease (Table 5). In two, the drug targeting FXI or FXIa was compared with a DOAC. In the PACIFIC-AF trial, Piccini et al compared asundexian (20 or 50 mg/d) with twice daily apixaban for patients ≥ 45 years of age with nonvalvular atrial fibrillation and at least 1 bleeding risk factor (bleeding within past 12 months, glomerular filtration rate 30-50 mL/min, or current indication for aspirin).⁸⁷ Ratios of incidence for the primary end point, major or clinically relevant nonmajor (CRNM) bleeding over 12 weeks, were significantly lower in patients on asundexian 20 mg

Table 3. Trials of drugs targeting FXIa for secondary prevention of ischemic stroke

Clinical trial	Study drug	Patients	Primary end point	Patient population	Incidence of bleeding
PACIFIC-STROKE	Asundexian (10, 20, or 50 mg/d) compared with placebo	More than 45 y with acute noncardioembolic stroke	Symptomatic ischemic stroke or MRI-detected covert Infarct at 26 wks	Asundexian 10 mg (19%)* Asundexian 20 mg (22%)* Asundexian 50 mg (20%)* Placebo (19%)	Major or CRNM bleeding (ISTH) Asundexian 10 mg (4%) Asundexian 20 mg (3%) Asundexian 50 mg (4%) Placebo (2%)
AXIOMATIC-SSP	Milvexian (25 to 200 mg once or twice daily) compared with placebo	More than 40 y with acute ischemic stroke (<48 h) or high-risk TIA	Symptomatic ischemic stroke or MRI-detected covert Infarct at 90 weeks	Twice daily dosing: Milvexian 25 mg (18.5%)* Milvexian 50 mg (14.1%)* Milvexian 100 mg (14.8%)* Milvexian 200 mg (16.4%)* Placebo (16.1%)	BARC-modified 3 or 5 bleeding Milvexian 25 mg (0.6%) Milvexian 50 mg (1.5%) Milvexian 100 mg (1.6%) Milvexian 200 mg (1.5%) Placebo (1%)

BARC, Bleeding Academic Research Consortium; CRNM, clinically relevant nonmajor bleeding; TIA, transient ischemic attack

*Not significantly different from results with placebo.

Table 4. Phase 3 studies involving asundexian, milvexian, and abelacimab

Study drug	Trial	Comparator	Estimated enrollment	Patients and primary outcome
Asundexian	OCEANIC-STROKE	Placebo	9 300	Patients: acute noncardioembolic ischemic stroke or high-risk transient ischemic attack Efficacy: time to first occurrence of ischemic stroke Safety: time to first occurrence of ISTH major bleeding
	OCEANIC-AF	Apixaban	18 000	Patients: atrial fibrillation at risk of stroke Efficacy: time to first occurrence of stroke or systemic embolism Safety: time to first occurrence of ISTH major bleeding
Milvexian	LIBREXIA-ACS	Placebo	16 000	Patients: acute coronary syndrome, with percutaneous intervention or conservative treatment, on antiplatelet agents Efficacy: time to occurrence of ischemic stroke
	LIBREXIA-STROKE	Placebo	15 000	Patients: acute stroke or high-risk transient ischemic attack on antiplatelet therapy Efficacy: time to occurrence of major adverse cardiovascular event (death, MI, or stroke)
	LIBREXIA-AF	Apixaban	15 500	Patients: atrial fibrillation Efficacy: time to first occurrence of stroke or non-central nervous system embolization
Abelacimab	LILAC-TIMI 76	Placebo	1 900	Patients: patients who are at high risk with atrial fibrillation deemed unsuitable for oral anticoagulation Efficacy: time to first ischemic stroke or systemic embolism Safety: time to first occurrence of BARC type 3c/5 bleeding
	ASTER	Apixaban	1 655	Patients: patients with cancer with venous thromboembolism Efficacy: time to VTE recurrence (new deep vein thrombosis, new pulmonary embolism, or fatal pulmonary embolism)
	MAGNOLIA	Dalteparin	1 020	Patients: gastrointestinal and genitourinary cancers with venous thromboembolism Efficacy: time to VTE recurrence (new deep vein thrombosis, new pulmonary embolism, or fatal pulmonary embolism)

(0.50; 90% CI, 0.14-1.68) or 50 mg (0.16; 90% CI, 0.01-0.99) than in those on apixaban. PACIFIC-AF was the first trial to show that patients on a FXIa inhibitor bleed less than those on a DOAC. The study had limitations, including a brief follow-up period (12 weeks), a small number of bleeding events

(10 episodes across all arms), and no major bleeding (all events were CRNM bleeds).

The AZALEA-TIMI 71 (ANT-006) trial compared patients treated with the anti-FXI IgG abelacimab (90 or 150 mg subcutaneously

Table 5. Clinical trials of drugs targeting FXI or FXIa for patients with heart disease

Clinical trial	Study drug	Comparator	Primary end point	Patient population	Incidence of bleeding
PACIFIC-AF	Asundexian	Apixaban	Major or CNMR bleeding (ISTH) out to 12 wk	At least 45 y nonvalvular atrial fibrillation, CHAD2DS2-VASc score ≥ 2 if male and ≥ 3 if female.	Asundexian 20 mg (1.2%)* Asundexian 50 mg (0.4%)* Apixaban 5 mg BID (2.4%)
PACIFIC-AMI	Asundexian	Placebo	BARC level 2, 3, or 5 bleeding out to 6 mo	At least 45 y, acute MI by clinical symptoms and elevated myocardial necrosis markers.	Asundexian 10 mg (7.6%)† Asundexian 20 mg (8.1%)† Asundexian 50 mg (10.5%)† Placebo (9.0%)
AZALEA-TIMI 71	Abelacimab	Rivaroxaban	Major or CNMR bleeding (ISTH) (average follow-up, 21 mo)	At least 55 y, nonvalvular atrial fibrillation, and a moderate-to-high risk of embolic stroke.	Abelacimab 90 mg (1.9/100 patient years)‡ Abelacimab 150 mg (2.7/100 patient years)‡ Rivaroxaban 20 mg/d (8.1/100 patient years)

BARC, Bleeding Academic Research Consortium; BID, twice daily; CHAD2DS2-VASc, a scoring system that stratifies patients with stroke risk and atrial fibrillation. The acronym stands for congestive heart failure, hypertension, age ≥ 75 years (doubled), diabetes, stroke (doubled), vascular disease, age between 65 and 74 years, and sex (female); ISTH, International Society on Thrombosis and Haemostasis.

*Significantly lower than apixaban 5 mg twice a day.

†Not significantly different from placebo.

‡Significantly lower than rivaroxaban 20 mg/d ($P < .0001$).

monthly) with those treated with rivaroxaban (20 mg/d; 15 mg if GFR \leq 50 mL/min).⁸⁸ Patients were followed for ~2 years, providing information on long-term treatment. Enrollees were aged \geq 55 years with atrial fibrillation at moderate-to-high risk for embolic stroke. The primary end point was International Society on Thrombosis and Haemostasis major or CRNM bleeding. Median free FXI antigen was reduced by 97% (interquartile range [IQR], 50%-99%) and 99% (IQR, 98%-99%) over the dosing interval in patients on 90 and 150 mg abelacimab, respectively, indicating a prolonged state of severe FXI deficiency in most patients.

AZALEA-TIMI 71 was stopped early because reductions in bleeding for patients on abelacimab, compared with rivaroxaban, substantially exceeded expectation. Incidences for the primary end point in patients on 90 and 150 mg abelacimab were 1.9 and 2.7 cases per 100 patient years, respectively, compared with 8.1 per 100 patient years with rivaroxaban. Over a median follow-up of 20.7 months, the primary end point was reduced by 67% (hazard ratio [HR], 0.33; 95% CI, 0.19-0.55; $P < .0001$) in patients on 150 mg abelacimab compared with those on rivaroxaban, with a 74% reduction in major bleeding (HR 0.26; 95% CI, 0.11-0.61; $P < .002$), and a 93% reduction in gastrointestinal bleeding (HR 0.07; 95% CI, 0.01-0.50; $P < .008$). Intracranial bleeds were rare and comparable for 150 mg abelacimab and rivaroxaban (0.3 and 0.6 cases per 100 patient years, respectively). AZALEA-TIMI 71 is the first study to show reduced major bleeding with a FXI/XIa inhibitor compared with a DOAC, using drug doses previously shown to reduce VTE in patients undergoing TKA.⁷⁵

In the PACIFIC-AMI trial, Rao et al compared asundexian (10, 20, or 50 mg/d) with placebo for patients with acute MI on antiplatelet therapy.⁸⁹ The primary outcome (Bleeding Academic Research Consortium level 2, 3, or 5 bleeding) occurred at similar rates in patients on asundexian (7.6% to 10.5%) and placebo (9.0%) over a median follow-up of 368 days. Data from the PACIFIC-STROKE and AXIOMATIC-SSP trials had suggested that superimposing FXIa inhibition on antiplatelet therapy was safe,^{85,86} and the PACIFIC-AMI trial supports this impression with data collected over a longer period.

FXI or FXIa inhibition in patients with renal failure

FXI/XIa inhibitors are undergoing testing for patients at increased risk of thromboembolism who are not good candidates for treatment with a DOAC or VKA. Patients with chronic renal failure are not only at increased risk of MI, stroke, and VTE^{90,91} but also at high risk of bleeding.^{90,92} Lorentz et al compared single doses of the anti-FXI IgG xisomab (0.25 or 0.50 mg/kg) with placebo for patients undergoing heparin-free hemodialysis.⁷⁶ Xisomab reduced occlusive events requiring circuit dialysis exchange by ~60% compared with occlusion rates in the same patients before receiving xisomab. There was no evidence of compromised hemostasis. Xisomab also lowered plasma C-reactive protein and thrombin-antithrombin complex levels. Phase 2 trials assessing the safety of Ionis-FXI Rx (EMERALD), the new generation ASO BAY2976217

(Re-THINC), and osocimab (CONVERT) in adults with end-stage renal disease on hemodialysis are underway.⁹⁰

Summary of clinical studies

Phase 2 studies of FXI/XIa inhibitors provide preliminary indications of efficacy and safety. At this point, evidence for efficacy is limited to demonstrations that FXI or FXIa inhibition prevents VTE in patients undergoing TKA^{74,75,77,84} and reduces clotting in hemodialysis circuits.⁷⁶ Superimposing FXIa inhibition on antiplatelet therapy did not reduce recurrent stroke but might have reduced recurrent symptomatic ischemic stroke. The effectiveness of FXI or FXIa inhibition relative to DOACs remains to be established. The phase 3 OCEANIC-AF, LIBREXIA-AF, and ASTER trials (Table 4) are designed to address this issue. On 19 November 2023, the OCEANIC-AF trial comparing asundexian with apixaban for prevention of stroke or embolization in patients with atrial fibrillation was stopped because data surveillance indicated inferior efficacy for asundexian.⁹³ This raises the possibility that FXI/XIa inhibitors are not as effective as DOACs for reducing thromboembolism associated with atrial fibrillation. Unlike the other drugs listed in table 4, asundexian was not tested as prophylaxis in knee replacement surgery, which would have facilitated comparison to other FXI/XIa targeting drugs. Several doses of asundexian were tested for recurrent stroke prevention,⁸⁵ but none affected the primary study outcome. At this point, therefore, we lack the information needed to determine if the OCEANIC-AF results reflect a limitation of FXI/XIa inhibitors as a class, or a limitation specific to asundexian at the doses tested.

In head-to-head comparisons in the PACIFIC-AF and AZALEA-TIMI 71 trials, FXI/XIa inhibitors caused less bleeding than DOACs. The results of the AZALEA-TIMI 71 trial are worth emphasizing. In this trial, a high dose of an antibody was used to essentially ablate FXI for >20 months. The profound reduction in overall major and CRNM bleeding when compared with rivaroxaban, and the very low incidence of intracranial and gastrointestinal bleeding supports the premise that FXI or FXIa inhibition will have a significant safety advantage over currently used anticoagulants and that long-term near total reduction of FXI or FXIa activity does not place patients at unacceptable risk of severe bleeding.

Future considerations, FXI or FXIa inhibition and inflammation

The FXI gene arose from a duplication of the gene for prekallikrein, the precursor of the protease plasma kallikrein.⁹⁴ Plasma kallikrein contributes to inflammation and kinin formation, and FXI retains some of the functional characteristics of its parent molecule.^{24,25} FXI contributes to the pathology of sepsis in primate and rodent models.⁹⁵⁻⁹⁹ Although this may partly reflect its role in thrombin generation, FXI also supports the systemic inflammatory response to infection. FXI deficiency or treatment with xisomab reduced the early "cytokine storm" in sepsis models. Curiously, although FXI is a downstream substrate for FXIIa during contact activation (Figure 1B), FXI inhibition blunted FXII activation in vivo during sepsis.^{98,99} These data suggest that FXI, rather than serving as a link in a unidirectional chain of enzymatic reactions that culminates in

thrombin generation, resides at a nexus where several host-defense mechanisms converge.

Conclusions

In 2005, Hirsh et al proposed that an ideal anticoagulant should have a high efficacy-to-safety index and a predictable dose response so that laboratory monitoring is not required.¹⁰⁰ DOACs are improvements on VKAs in both regards. FXI or FXIa inhibitors may further uncouple antithrombotic effects from adverse effects on hemostasis.³ If phase 3 trials demonstrate that these drugs combine efficacy with improved safety, then antithrombotic therapy may become available for many patients who currently go untreated because of bleeding concerns.

Acknowledgments

The authors acknowledge generous support from the National Institutes of Health, National Heart, Lung, and Blood Institute awards R35 HL140025, HL101972, and HL144113 and the Ernest W. Goodpasture Chair in Experimental Pathology for Translational Research, Vanderbilt University Medical Center.

Authorship

Contribution: D.G. and A.G. contributed to writing the manuscript.

Conflict-of-interest disclosure: D.G. is a consultant for pharmaceutical companies (Anthos Therapeutics; Aronora, Inc; Bayer Pharma; Bristol Myers Squibb; Ionis Pharmaceuticals; and Janssen Pharmaceuticals), with interests in targeting factor XI, factor XII, and prekallikrein for therapeutic purposes. A.G. is an employee of Aronora Inc, with interests in developing safe antithrombotic agents.

ORCID profiles: D.G., 0000-0001-8142-8014; A.G., 0000-0001-6212-2247.

Correspondence: David Gailani, Pathology, Microbiology and Immunology, Vanderbilt University Medical Center, Room 4918, The Vanderbilt Clinic, 1301 Medical Center Dr, Nashville, TN 37232; email: dave.gailani@vanderbilt.edu; and Andras Gruber, Aronora, Inc, 1818 SW Ave, Suite 102, Portland, OR 97201; email: andras.gruber@aronorabio.com.

Footnote

Submitted 20 July 2023; accepted 7 December 2023; prepublished online on *Blood* First Edition 24 December 2023. <https://doi.org/10.1182/blood.2023020722>.

REFERENCES

- Macfarlane RG. Hemostasis: introduction. *Br Med Bull*. 1977;33(3):183-185.
- Macfarlane RG. An enzyme cascade in the blood clotting mechanism, and its function as a biochemical amplifier. *Nature*. 1964; 202:498-499.
- Hsu C, Hutt E, Bloomfield DM, Gailani D, Weitz JI. Factor XI inhibition to uncouple thrombosis from hemostasis: JACC review topic of the week. *J Am Coll Cardiol*. 2021; 78(6):625-631.
- Fredenburgh JC, Weitz JI. News at XI: moving beyond factor Xa inhibitors. *J Thromb Haemost*. 2023;21(7):1692-1702.
- Negrier C, Shima M, Hoffman M. The central role of thrombin in bleeding disorders. *Blood Rev*. 2019;38:100582.
- Brummel-Ziedins K, Mann KG, Fredenburgh JC, Weitz JI. Molecular basis of blood coagulation. In: Hoffman R, Benz EJ, Silberstein LE, et al., eds. *Hematology: Basic Principles and Practices*. 8th ed. Elsevier; 2023:1968-1987.
- Repke D, Gemmell CH, Guha A, Turitto VT, Broze GJ Jr, Nemerson Y. Hemophilia as a defect of the tissue factor pathway of blood coagulation: effect of factors VIII and IX on factor X activation in a continuous-flow reactor. *Proc Natl Acad Sci U S A*. 1990; 87(19):7623-7627.
- Roberts HR, Monroe DM, Oliver JA, Chang JY, Hoffman M. Newer concepts of blood coagulation. *Haemophilia*. 1998;4: 331-334.
- Mackman N. Role of tissue factor in hemostasis, thrombosis and vascular development. *Arterioscler Thromb Vasc Biol*. 2004;24(6):1015-1022.
- Smith SA, Travers RJ, Morrissey JH. How it all starts: initiation of the clotting cascade. *Crit Rev Biochem Mol Biol*. 2015;50(4): 326-336.
- McManus MP, Gailani D. Mouse models of coagulation factor deficiencies. In: Wang X, ed. *Animal Models of Diseases: Translational Medicine Perspective for Drug Discovery and Development*. Bentham Scientific Publishers; 2012:67-121.
- Ruff CT, Giugliano RP, Braunwald E, et al. Comparison of the efficacy and safety of new oral anticoagulants with warfarin in patients with atrial fibrillation: a meta-analysis of randomised trials. *Lancet*. 2014; 383(9921):955-962.
- van Es N, Coppens M, Schulman S, Middeldorp S, Buller HR. Direct oral anticoagulants compared with vitamin K antagonists for acute venous thromboembolism: evidence from phase 3 trials. *Blood*. 2014;124(12):1968-1975.
- Palareti G. Direct oral anticoagulants and bleeding risk (in comparison to vitamin K antagonists and heparins), and the treatment of bleeding. *Semin Hematol*. 2014;51(2):102-111.
- Al-Khatib SM, Pokorney SD, Al-Khalidi HR, et al. Underuse of oral anticoagulants in privately insured patients with atrial fibrillation: a population being targeted by the implementation of a randomized controlled trial to improve treatment with oral anticoagulants in patients with atrial fibrillation (IMPACT-AFib). *Am Heart J*. 2020;229:110-117.
- Park S, Je NK. Underutilization of anticoagulants in patients with nonvalvular atrial fibrillation in the era of non-vitamin K antagonist oral anticoagulants. *Int J Arrhythm*. 2022;23:1.
- Sanghai S, Wong C, Wang Z, et al. Rates of potentially inappropriate dosing of direct-acting oral anticoagulants and associations with geriatric conditions among older patients with atrial fibrillation: the SAGE-AF study. *J Am Heart Assoc*. 2020;9(6): e014108.
- Khan F, Tritschler T, Kimpton M, et al. Long-term risk for major bleeding during extended oral anticoagulant therapy for first unprovoked venous thromboembolism: a systematic review and meta-analysis. *Ann Intern Med*. 2021;174(10):1420-1429.
- Li S, Zhang Y. Do oral factor Xa inhibitors have a role in patients with mechanical heart valves? *Am J Cardiovasc Drugs*. 2022;22(6): 595-600.
- Eikelboom JW, Weitz JI. Warfarin faring better: vitamin K antagonists beat rivaroxaban and apixaban in the INVICTUS and PROACT Xa trials. *J Thromb Haemost*. 2023;21(11):3067-3071.
- Pastori D, Menichelli D, Cammisotto V, Pignatelli P. Use of direct oral anticoagulants in patients with antiphospholipid syndrome: a systematic review and comparison of the international guidelines. *Front Cardiovasc Med*. 2021;8:715878.
- Wu X, Cao S, Yu B, He T. Comparing the efficacy and safety of direct oral anticoagulants versus Vitamin K antagonists in patients with antiphospholipid syndrome: a systematic review and meta-analysis. *Blood Coagul Fibrinolysis*. 2022;33(7): 389-401.
- Lip GYH. Anticoagulation in atrial fibrillation and rheumatic heart disease. *N Engl J Med*. 2022;387(11):1036-1038.
- Mohammed BM, Matafonov A, Ivanov I, et al. An update on factor XI structure and function. *Thromb Res*. 2018;161:94-105.
- Moellmer SA, Puy C, McCarty OJT. Biology of factor XI. *Blood*. 2024;143(15):1445-1454.
- Rosenthal RL, Dreskin OH, Rosenthal N. Plasma thromboplastin antecedent (PTA)

- deficiency: clinical, coagulation, therapeutic and hereditary aspects of a new hemophilia-like disease. *Blood*. 1955;10(2):120-131.
27. Konrath S, Mailer RK, Renné T. Mechanism, functions, and diagnostic relevance of FXII activation by foreign surfaces. *Hamostaseologie*. 2021;41(6):489-501.
28. Gailani D, Tillman BF, Wheeler AP. Rare coagulation factor deficiencies. In: molecular basis of blood coagulation. In: Hoffman R, Benz EJ, Silberstein LE, et al., eds. *Hematology: Basic Principles and Practices*. 8th ed. Elsevier; 2023:2125-2143.
29. Barg AA, Livnat T, Kenet G. Factor XI deficiency: phenotypic age-related considerations and clinical approach towards bleeding risk assessment. *Blood*. 2024;143(15):1455-1464.
30. Lewandowska MD, Connors JM. Factor XI deficiency. *Hematol Oncol Clin North Am*. 2021;35(6):1157-1169.
31. Maas DPMSM, Saes JL, Blijlevens NMA, et al. Treatment of patients with rare bleeding disorders in the Netherlands: real-life data from the RBiN study. *J Thromb Haemost*. 2022;20(4):833-844.
32. Ragni MV, Sinha D, Seaman F, Lewis JH, Spero JA, Walsh PN. Comparison of bleeding tendency, factor XI coagulant activity, and factor XI antigen in 25 factor XI-deficient kindreds. *Blood*. 1985;65(3):719-724.
33. Bolton-Maggs PH. Factor XI deficiency, resolving the enigma. *Hematology Am Soc Hematol Educ Program*. 2009;2009(1):97-105.
34. Naito K, Fujikawa K. Activation of human blood coagulation factor XI independent of factor XII. Factor XI is activated by thrombin and factor XIa in the presence of negatively charged surfaces. *J Biol Chem*. 1991;266(12):7353-7358.
35. Gailani D, Broze GJ Jr. Factor XI activation in a revised model of blood coagulation. *Science*. 1991;253(5022):909-912.
36. Von dem Borne PA, Bajzar L, Meijers JC, Nesheim ME, Bouma BN. Thrombin-mediated activation of factor XI results in a thrombin-activatable fibrinolysis inhibitor-dependent inhibition of fibrinolysis. *J Clin Invest*. 1997;99(10):2323-2327.
37. Gruber A, Hanson SR. Factor XI-dependence of surface- and tissue factor-initiated thrombus propagation in primates. *Blood*. 2003;102(3):953-955.
38. Tucker EI, Marzec UM, White TC, et al. Prevention of vascular graft occlusion and thrombus-associated thrombin generation by inhibition of factor XI. *Blood*. 2009;113(4):936-944.
39. Cheng Q, Tucker EI, Pine MS, et al. A role for factor XIIa-mediated factor XI activation in thrombus formation in vivo. *Blood*. 2010;116(19):3981-3989.
40. Rosen ED, Gailani D, Castellino FJ. FXI is essential for thrombus formation following FeCl₃-induced injury of the carotid artery in the mouse. *Thromb Haemost*. 2002;87(4):774-776.
41. Wang X, Cheng Q, Xu L, et al. Effects of factor IX or factor XI deficiency on ferric chloride-induced carotid artery occlusion in mice. *J Thromb Haemost*. 2005;3(4):695-702.
42. Leung PY, Hurst S, Berny-Lang MA, et al. Inhibition of factor XII-mediated activation of factor XI provides protection against experimental acute ischemic stroke in mice. *Transl Stroke Res*. 2012;3:381-389.
43. van Montfoort ML, Knaup VL, Marquart JA, et al. Two novel inhibitory anti-human factor XI antibodies prevent cessation of blood flow in a murine venous thrombosis model. *Thromb Haemost*. 2013;110(5):1065-1073.
44. Lorentz CU, Verbout NG, Cao Z, et al. Factor XI contributes to myocardial ischemia-reperfusion injury in mice. *Blood Adv*. 2018;2:85-88.
45. Gui T, Reheman A, Funkhouser WK, et al. In vivo response to vascular injury in the absence of factor IX: examination in factor IX knockout mice. *Thromb Res*. 2007;121(2):225-234.
46. Renné T, Pozgajová M, Grüner S, et al. Defective thrombus formation in mice lacking coagulation factor XII. *J Exp Med*. 2005;202(2):271-281.
47. Kleinschnitz C, Stoll G, Bendszus M, et al. Targeting coagulation factor XII provides protection from pathological thrombosis in cerebral ischemia without interfering with hemostasis. *J Exp Med*. 2006;203(3):513-518.
48. Gailani D, Gruber A. Factor XI as a therapeutic target. *Arterioscler Thromb Vasc Biol*. 2016;36(7):1316-1322.
49. Matafonov A, Leung PY, Gailani AE, et al. Factor XII inhibition reduces thrombus formation in a primate thrombosis model. *Blood*. 2014;123(11):1739-1746.
50. Meijers JC, Tekelenburg WL, Bouma BN, Bertina RM, Rosendaal FR. High levels of coagulation factor XI as a risk factor for venous thrombosis. *N Engl J Med*. 2000;342(10):696-701.
51. Bezemer ID, Bare LA, Doggen CJ, et al. Gene variants associated with deep vein thrombosis. *JAMA*. 2008;299(11):1306-1314.
52. Li Y, Bezemer ID, Rowland CM, et al. Genetic variants associated with deep vein thrombosis: the F11 locus. *J Thromb Haemost*. 2009;7(11):1802-1808.
53. Bruzelius M, Ljungqvist M, Bottai M, et al. F11 is associated with recurrent VTE in women. A prospective cohort study. *Thromb Haemost*. 2016;115(2):406-414.
54. Preis M, Hirsch J, Kotler A, et al. Factor XI deficiency is associated with lower risk for cardiovascular and venous thromboembolism events. *Blood*. 2017;129(9):1210-1215.
55. Cushman M, O'Meara ES, Folsom AR, Heckbert SR. Coagulation factors IX through XIII and the risk of future venous thrombosis: the Longitudinal Investigation of Thromboembolism Etiology. *Blood*. 2009;114(14):2878-2883.
56. Berliner JL, Rybicki AC, Kaplan RC, Monrad ES, Freeman R, Billett HH. Elevated levels of Factor XI are associated with cardiovascular disease in women. *Thromb Res*. 2002;107(1-2):55-60.
57. Doggen CJ, Rosendaal FR, Meijers JC. Levels of intrinsic coagulation factors and the risk of myocardial infarction among men: opposite and synergistic effects of factors XI and XII. *Blood*. 2006;108(13):4045-4051.
58. Siegerink B, Rosendaal FR, Algra A. Antigen levels of coagulation factor XII, coagulation factor XI and prekallikrein, and the risk of myocardial infarction and ischemic stroke in young women. *J Thromb Haemost*. 2014;12(5):606-613.
59. Yang DT, Flanders MM, Kim H, Rodgers GM. Elevated factor XI activity levels are associated with an increased odds ratio for cerebrovascular events. *Am J Clin Pathol*. 2006;126(3):411-415.
60. Suri MF, Yamagishi K, Aleksic N, Hannan PJ, Folsom AR. Novel hemostatic factor levels and risk of ischemic stroke: the Atherosclerosis Risk in Communities (ARIC) study. *Cerebrovasc Dis*. 2010;29(5):497-502.
61. Salomon O, Steinberg DM, Zucker M, Varon D, Zivelin A, Seligsohn U. Patients with severe factor XI deficiency have a reduced incidence of deep-vein thrombosis. *Thromb Haemost*. 2011;105(2):269-273.
62. Salomon O, Steinberg DM, Koren-Morag N, Tanne D, Seligsohn U. Reduced incidence of ischemic stroke in patients with severe factor XI deficiency. *Blood*. 2008;111(8):4113-4117.
63. Lämmle B, Willemin WA, Huber I, et al. Thromboembolism and bleeding tendency in congenital factor XII deficiency—a study on 74 subjects from 14 Swiss families. *Thromb Haemost*. 1991;65(2):117-121.
64. Endler G, Marsik C, Jilma B, Schickbauer T, Quehenberger P, Mannhalter C. Evidence of a U-shaped association between factor XII activity and overall survival. *J Thromb Haemost*. 2007;5(6):1143-1148.
65. Kravtsov DV, Matafonov A, Tucker EI, et al. Factor XI contributes to thrombin generation in the absence of factor XII. *Blood*. 2009;114(2):452-458.
66. Tillman B, Gailani D. Inhibition of factors XI and XII for prevention of thrombosis induced by artificial surfaces. *Semin Thromb Hemost*. 2018;44(1):60-69.
67. Jaffer IH, Weitz JI. The blood compatibility challenge. Part 1: blood-contacting medical devices: the scope of the problem. *Acta Biomater*. 2019;94:2-10.
68. Bartlett R, Arachichilage DJ, Chitlur M, et al. The history of extracorporeal membrane oxygenation and the development of

- extracorporeal membrane oxygenation anticoagulation. *Semin Thromb Hemost*. 2024;50(1):81-90.
69. Yau JW, Liao P, Fredenburgh JC, et al. Selective depletion of factor XI or factor XII with antisense oligonucleotides attenuates catheter thrombosis in rabbits. *Blood*. 2014;123(13):2102-2107.
 70. Litvak M, Shamanaev A, Zalawadiya S, et al. Titanium is a potent inducer of contact activation: implications for intravascular device. *J Thromb Haemost*. 2023;21(5):1200-1213.
 71. de Jong CMM, Blondon M, Ay C, et al. Incidence and impact of anticoagulation-associated abnormal menstrual bleeding in women after venous thromboembolism. *Blood*. 2022;140(16):1764-1773.
 72. Bolton-Maggs PH, Patterson DA, Wensley RT, Tuddenham EG. Definition of the bleeding tendency in factor XI-deficient kindreds—a clinical and laboratory study. *Thromb Haemost*. 1995;73(2):194-202.
 73. Crosby JR, Marzec U, Revenko AS, et al. Antithrombotic effect of antisense factor XI oligonucleotide treatment in primates. *Arterioscler Thromb Vasc Biol*. 2013;33(7):1670-1678.
 74. Büller HR, Bethune C, Bhanot S, et al. Factor XI antisense oligonucleotide for prevention of venous thrombosis. *N Engl J Med*. 2015;372(3):232-240.
 75. Verhamme P, Yi BA, Segers A, et al. ANT-005 TKA investigators. Abrelcimab for prevention of venous thromboembolism. *N Engl J Med*. 2021;385(7):609-617.
 76. Lorentz CU, Tucker EI, Verbout NG, et al. The contact activation inhibitor AB023 in heparin-free hemodialysis: results of a randomized phase 2 clinical trial. *Blood*. 2021;138(22):2173-2184.
 77. Weitz JI, Bauersachs R, Becker B, et al. Effect of osocimab in preventing venous thromboembolism among patients undergoing knee arthroplasty: the FOXTROT randomized clinical trial. *JAMA*. 2020;323(2):130-139.
 78. Dilger AK, Pabbisetty KB, Corte JR, et al. Discovery of milvexian, a high-affinity, orally bioavailable inhibitor of factor XIa in clinical studies for antithrombotic therapy. *J Med Chem*. 2022;65(3):1770-1785.
 79. Heitmeier S, Visser M, Tersteegen A, et al. Pharmacological profile of asundexian, a novel, orally bioavailable inhibitor of factor XIa. *J Thromb Haemost*. 2022;20(6):1400-1411.
 80. Harrington J, Piccini JP, Alexander JH, Granger CB, Patel MR. Clinical evaluation of factor XIa inhibitor drugs: JACC review topic of the week. *J Am Coll Cardiol*. 2023;81(8):771-779.
 81. Greco A, Laudani C, Spagnolo M, et al. Capodanno D pharmacology and clinical development of factor XI inhibitors. *Circulation*. 2023;147(11):897-913.
 82. Bentounes NK, Melicine S, Martin AC, Smadja DM, Gendron N. Development of new anticoagulant in 2023: prime time for anti-factor XI and XIa inhibitors. *J Med Vasc*. 2023;48(2):69-80.
 83. Chan NC, Weitz JI. New therapeutic targets for the prevention and treatment of venous thromboembolism with a focus on factor XI inhibitors. *Arterioscler Thromb Vasc Biol*. 2023;43(10):1755-1763.
 84. Weitz JI, Strony J, Ageno W, et al. Milvexian for the prevention of venous thromboembolism. *N Engl J Med*. 2021;385(23):2161-2172.
 85. Shoamanesh A, Mundl H, Smith EE, et al. Factor XIa inhibition with asundexian after acute non-cardioembolic ischaemic stroke (PACIFIC-Stroke): an international, randomised, double-blind, placebo-controlled, phase 2b trial. *Lancet*. 2022;400(10357):997-1007.
 86. Sharma M, Molina CA, Toyoda K, et al. Safety and efficacy of factor XIa inhibition with milvexian for secondary stroke prevention (AXIOMATIC-SSP): a phase 2, international, randomised, double-blind, placebo-controlled, dose-finding trial. *Lancet Neurol*. 2024;23:46-59.
 87. Piccini JP, Caso V, Connolly SJ, et al; PACIFIC-AF Investigators. Safety of the oral factor XIa inhibitor asundexian compared with apixaban in patients with atrial fibrillation (PACIFIC-AF): a multicentre, randomised, double-blind, double-dummy, dose-finding phase 2 study. *Lancet*. 2022;399(10333):1383-1390.
 88. Ruff TR for the AZALEA-TIMI 71 Steering Committee. Abrelcimab, a novel factor XI/ XIa inhibitor versus rivaroxaban in patients with atrial fibrillation: primary results of the AZALEA-TIMI 71 randomized trial [late breaking abstract LBS.05 session]. American Heart Association Scientific Sessions; 2023.
 89. Rao SV, Kirsch B, Bhatt DL, et al. A multicenter, phase 2, randomized, placebo-controlled, double-blind, parallel-group, dose-finding trial of the oral factor XIa inhibitor asundexian to prevent adverse cardiovascular outcomes after acute myocardial infarction. *Circulation*. 2022;146(16):1196-1206.
 90. Eikelboom J, Floege J, Thadhani R, Weitz JI, Winkelmayer WC. Anticoagulation in patients with kidney failure on dialysis: factor XI as a therapeutic approach. *Kidney Int*. 2021;100(6):1199-1207.
 91. Go AS, Chertow GM, Fan D, McCulloch CE, Hsu CY. Chronic kidney disease and the risks of death, cardiovascular events, and hospitalization. *N Engl J Med*. 2004;351(13):1296-1305.
 92. Jalal DI, Chonchol M, Targher G. Disorders of hemostasis associated with chronic kidney disease. *Semin Thromb Hemost*. 2010;36(1):34-40.
 93. Bayer GlobalOCEANIC-AF study stopped early due to lack of efficacy. Accessed 19 November 2023. <https://www.bayer.com/media/en-us/oceanic-af-study-stopped-early-due-to-lack-of-efficacy/>
 94. Ponczek MB, Shamanaev A, LaPlace A, et al. The evolution of factor XI and the kallikrein-kinin system. *Blood Adv*. 2020;4(24):6135-6147.
 95. Tucker EI, Gailani D, Hurst S, Cheng Q, Hanson SR, Gruber A. Survival advantage of coagulation factor XI-deficient mice during peritoneal sepsis. *J Infect Dis*. 2008;198(2):271-274.
 96. Luo D, Szaba FM, Kummer LW, et al. Factor XI-deficient mice display reduced inflammation, coagulopathy, and bacterial growth during listeriosis. *Infect Immun*. 2012;80(1):91-99.
 97. Tucker EI, Verbout NG, Leung PY, et al. Inhibition of factor XI activation attenuates inflammation and coagulopathy while improving the survival of mouse polymicrobial sepsis. *Blood*. 2012;119(20):4762-4768.
 98. Bane CE Jr, Ivanov I, Matafonov A, et al. Factor XI deficiency alters the cytokine response and activation of contact proteases during polymicrobial sepsis in mice. *PLoS One*. 2016;11(4):e0152968.
 99. Silasi R, Keshari RS, Lupu C, et al. Inhibition of contact-mediated activation of factor XI protects baboons against *S aureus*-induced organ damage and death. *Blood Adv*. 2019;3(4):658-669.
 100. Hirsh J, O'Donnell M, Weitz JI. New anticoagulants. *Blood*. 2005;105(2):453-463.

© 2024 American Society of Hematology. Published by Elsevier Inc. All rights are reserved, including those for text and data mining, AI training, and similar technologies.